

BIOSYNTHESIS OF TROPIC ACID: FEEDING EXPERIMENTS WITH CINNAMOYL TROPINE AND LITTORINE

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Abstract—The administration of cinnamoyl-[2-¹⁴C]-tropine-[*N*-methyl-¹⁴C] to *Datura stramonium* plants resulted in the formation of labeled atropine and scopolamine. However the atropine was found to have almost all its radioactivity located on the *N*-methyl group of the alkaloid, indicating that the administered ester had undergone hydrolysis in the plant affording tropine and cinnamic acid, the latter not being utilized for the biosynthesis of tropic acid. Dual labeled *RS*-littorine (3 α -(2-hydroxy-3-phenylpropionyloxy)-[1-¹⁴C]-tropane-[3 β -³H]) was also fed to *D. stramonium* and radioactive atropine was obtained. However the drastic change in the ³H:¹⁴C ratio found in the atropine indicated that the littorine was not converted directly to the alkaloid, and it is suggested that the littorine is hydrolysed *in vivo* to tropine and phenyl-lactic acid, the latter undergoing rearrangement to tropic acid prior to esterification with tropine.

TROPIC ACID is found in nature as the acid moiety of the ester alkaloids atropine (7) and scopolamine. It was discovered in 1960¹ that the administration of phenylalanine-[3-¹⁴C] to intact *Datura stramonium* plants yielded radioactive atropine having essentially all its activity located at C-2 of its tropic acid moiety. Later workers confirmed this result in *D. stramonium* (intact plants)² and *D. metel* (sterile root culture).³ It was then established that the other carbons of the phenylalanine side chain are utilized for the production of the side chain of tropic acid.^{4,5} The pattern of labeling found in tropic acid after feeding variously labeled phenylalanines indicated that a migration of the carboxyl group from C-2 to C-3 occurs during the formation of tropic acid. By using phenylalanine-[U-¹⁴C] as a precursor it was established that the phenyl group is incorporated intact into tropic acid.⁶ The mechanism of the rearrangement of the side chain of phenylalanine is currently unknown. Various metabolites of phenylalanine have been examined as precursors of tropic acid. Phenylpyruvic acid (1) was incorporated, and in competition with phenylalanine it was almost as an efficient a precursor of tropic acid.⁷ However the expected facile interconversion of these two compounds (by transamination) does not enable us to make a decision

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¹ LEETE, E (1960) *J. Am. Chem. Soc.* **82**, 612

² UNDERHILL, E. W. and YOUNGKEN, H. W. (1962) *J. Pharm. Sci.* **51**, 121

³ GROSS, D. and SCHUTTE, H. R. (1963) *Arch. Pharm.* **296**, 1

⁴ LEETE, E. and LOUDEN, M. L. (1961) *Chem. Ind.* 1405

⁵ LOUDEN, M. L. and LEETE, E. (1962) *J. Am. Chem. Soc.* **84**, 1510, 4507

⁶ GIBSON, C. A. and YOUNGKEN, H. W. (1967) *J. Pharm. Sci.* **56**, 854

⁷ (a) LIEBISCH, H. W., BHAVSAR, G. C. and SCHALLER, H. J. (1969) *Biochem. Physiol. Alkaloide, Int. Symp.* (MOTHES, K., ed.), 4th Edn, p. 233, Akad., Berlin, E. Germany, (b) LIEBISCH, H. W. (1970) Abstracts of the 7th Int. Sympos. on the Chem. Natural Products, Riga, USSR, p. 557

as to which is biosynthetically closer to tropic acid. The same remarks apply to phenyl-lactic acid (**3**), the reduction product of phenylpyruvic acid. This acid is esterified with tropine in the alkaloid littorine (**5**), and phenylalanine was shown to be its precursor in *D. sanguinea*⁸ and *D. innoxia*.⁹ Phenyl-lactic acid is also a good precursor of tropic acid^{7,9} and it was reported to be superior to phenylalanine in competitive feeding experiments.

Cinnamic acid (**2**), a metabolite of phenylalanine in many plant species, was considered to be a prime candidate as an intermediate between phenylalanine and tropic acid.¹⁰ In a model system it has been shown that phenyl *trans*-3-phenylthiolglycidate (**4**) rearranges on treatment with boron trifluoride etherate to 2-formylphenylthiol acetate (**6**).^{11,12} This reaction involves a 1,2-migration of a thiol ester group, analogous to the migration of the carboxyl group which occurs during the biosynthesis of tropic acid *in vivo*. However cinnamic acid^{7b,9} or its epoxide¹³ failed to serve as precursors of tropic acid in *Datura* species.

It was considered that perhaps rearrangement of the C₆-C₃ skeleton occurs after esterification of tropine with an acid derived from phenylalanine. Cinnamoyl-tropine was thus prepared labeled with ¹⁴C at C-2 of its cinnamoyl moiety, and on the *N*-methyl group of the tropine base. This doubly labeled compound was fed to *D. stramonium* plants by the wick method. After 5 days the plants were harvested and yielded radioactive atropine and scopolamine. However hydrolysis of the alkaloids yielded tropic acid having low activity (see Experimental), the bulk of the activity residing in the tropine bases. Demethylation of the tropine obtained by hydrolysis of the atropine with phenyl chloroformate¹⁴ yielded inactive *N*-carbophenoxynortropine. These results are consistent with the cinnamoyl-tropine being hydrolysed in the plant to tropine and cinnamic acid, the latter not being converted to tropic acid. The incorporation of radioactivity into atropine and scopolamine based on the ¹⁴C present in the tropine moiety of the administered cinnamoyl-tropine was 0.94 and 0.91% respectively.

We then examined the ability of littorine to serve as a direct precursor of atropine. Phenyl-lactic-[1-¹⁴C] acid was prepared from glycine-[1-¹⁴C] by the sequence: glycine → *N*-acetylglycine → azlactone of *N*-acetyl- α -aminocinnamic acid → phenylpyruvic acid → phenyl-lactic acid using modifications of established procedures.¹⁵ This phenyl-lactic acid was esterified with tropine yielding littorine.¹⁶ This material was mixed with littorine prepared from tropine-[3 β -³H]¹⁴ to afford dual labeled littorine (³H·¹⁴C = 6.75) which was fed as before to *D. stramonium* plants. After 7 days the plants yielded radioactive atropine (³H·¹⁴C = 33). Hydrolysis of this atropine afforded tropine (labeled only with ³H) which on oxidation to tropinone lost all its radioactivity indicating that all the tritium was located at C-3. The tropic acid was degraded as previously described⁵ and was found to have essentially all its activity located on the carboxyl group. The drastic change in the ³H·¹⁴C ratio of the isolated atropine indicates that the phenyl-lactic acid moiety of littorine does not undergo a rearrangement to tropic acid whilst it is esterified with tropine. The specific labeling of the tropine and tropic acid with tritium and ¹⁴C respectively

⁸ EVANS, W. C. and WOOLLEY, V. A. (1969) *Phytochemistry* **8**, 2183.

⁹ EVANS, W. C., WOOLLEY, J. G. and WOOLLEY, V. A. ref. 7a p. 227.

¹⁰ SPENGLER, J. D. (1968) in *Comprehensive Biochemistry* (FLORENCE, M. and STOLT, E. H., eds.), Vol. 20, p. 294. Elsevier, Amsterdam.

¹¹ WEMPLER, J. N. (1970) *J. Am. Chem. Soc.* **92**, 6694.

¹² DOMAGALA, J. and WEMPLER, J. (1973) *Tetrahedron Letters* 1179.

¹³ LILIE, E. and BRAUNSTEIN, J. D. unpublished work.

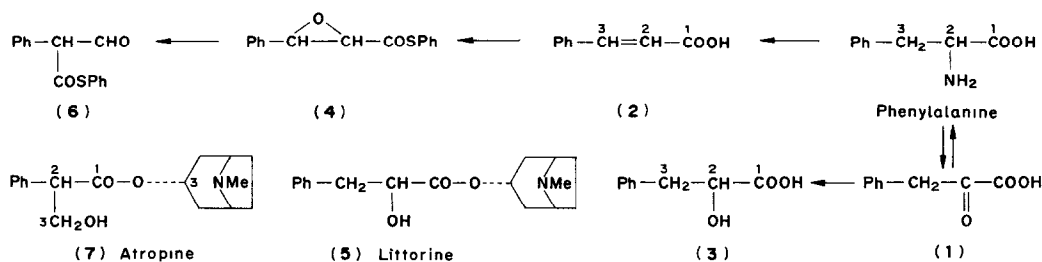
¹⁴ LILIE, E. (1972) *Phytochemistry* **11**, 1713.

¹⁵ HERBST, R. M. and SHEMIN, D. (1943) *Organic Syntheses* (BLATT, A. H., ed.), Coll. Vol. 2, pp. 1-11, 519.

¹⁶ TAKEUCHI, Y., KOGA, K., SUOJIBI, T. and YAMADA, S. (1971) *Chem. Pharm. Bull. (Japan)* **19**, 2603.

strongly suggests that the littorine is hydrolysed in the plant to tropine and phenyl-lactic acid, the latter undergoing rearrangement to tropic acid prior to esterification with tropine to yield atropine.

Our results could also be rationalized by postulating that the administered radioactive littorine undergoes a facile reversible hydrolysis in the plant, the resultant tropine and phenyl-lactic acid being then diluted with non-radioactive pools of these materials already present in the plant. It has been demonstrated that esters of tropane alcohols are readily hydrolysed in *Datura* and related species.¹⁷⁻¹⁹ The resynthesized littorine would then be expected to have a different $^3\text{H}:^{14}\text{C}$ ratio from the administered alkaloid. Rearrangement of the phenyl-lactic acid moiety of the littorine could then yield atropine having a $^3\text{H}:^{14}\text{C}$ ratio different from the administered littorine. This explanation seems unlikely since the littorine isolated from the plant at the end of the feeding experiment had essentially the same specific activity and $^3\text{H}:^{14}\text{C}$ ratio (6.9) as the administered alkaloid



Hypothetical metabolism of phenylalanine in *Datura*

EXPERIMENTAL

General methods Radioactivity measurements were carried out in a Nuclear Chicago Mark II liquid scintillation counter. Radioactive compounds were dissolved in 1–2 ml EtOH or H₂O and diluted with 10 ml dioxane soln containing naphthalene (10%), 2,5-diphenyloxazole (0.7%), and 1,4-bis-2-(5-phenyloxazolyl)benzene (0.05%).

Cinnamoyl-[2- ^{14}C]-tropine-[N-methyl- ^{14}C] Cinnamic-[2- ^{14}C] acid (ICN, Irvine, Calif, nominal activity 0.2 mCi, 85 mg 0.575 mM) dissolved in light petrol (10 ml) was refluxed with thionyl chloride (5 ml) for 3 hr. The reaction mixture was evaporated, the residue dissolved in petrol and the evaporation repeated. Tropine (100 mg, 0.71 mM) was dissolved in dry ether and the soln saturated with HCl gas. Evaporation yielded tropine hydrochloride which was mixed with the cinnamoyl chloride and the mixture heated at 100° for 6 hr. After cooling, the reaction mixture was dissolved in 2 N H₂SO₄, filtered and the filtrate made basic with Na₂CO₃ when cinnamoyl-[2- ^{14}C]-tropine monohydrate separated (115 mg, 7.8×10^8 dpm/mM) m.p. 45° (lit.²⁰ m.p. 45–46°). Cinnamoyl-tropine-[N-methyl- ^{14}C] was similarly prepared using cinnamoyl chloride and tropine-[N-methyl- ^{14}C].¹⁴

Administration of cinnamoyl-tropine to *D. stramonium* plants and isolation of the alkaloids For administration to the plants a mixture of cinnamoyl-[2- ^{14}C]-tropine (24.3 mg, 6.57×10^7 dpm) and cinnamoyl-tropine-[N-methyl- ^{14}C] (21.1 mg, 8.75×10^7 dpm) was dissolved in H₂O (6 ml) containing 0.1 ml 2 N H₂SO₄. This soln was administered to 7 *D. stramonium* plants (3-months-old) by means of cotton wicks inserted into the stems near to ground level. After 5 days the whole plants (fr. wt 250 g) were extracted as previously described,¹ inactive atropine (1 mM) and scopolamine (1 mM) being added as carriers to facilitate the isolation of the labeled alkaloids. The crude alkaloids had an activity of 2.3×10^7 dpm (15% of the activity fed to the plants). TLC on a sample of this mixture (on Silica gel G, developing with CHCl₃-EtOH-conc. NH₃, 85:14:1) showed the presence of the following alkaloids having the indicated distribution of radioactivity: atropine (R_f 0.3) (15.5%), cinnamoyl-tropine (R_f 0.72) (71%), and scopolamine (R_f 0.85) (13.6%). The alkaloids were separated by partition chromatography on a celite column,¹ atropine (8.2×10^5 dpm/mM) and scopolamine (8.0×10^5 dpm/mM) being obtained.

¹⁷ NEUMANN, D. and TSCHOPE, K. H. (1966) *Flora (Jena)* **156**, 521.

¹⁸ KACZKOWSKI, J. (1966). *Abh. Dtsch. Akad. Wiss. Berlin., Kl. Chem. Geol. Biol. Nr.* **3**, 521.

¹⁹ ACHARI, R., EVANS, W. C. and NEWCOMBE, F. (1969) *Naturwissenschaften* **56**, 88.

²⁰ JOWETT, H. A. D. and PYMAN, F. L. (1909) *J. Chem. Soc.* **95**, 1020.

Degradation of the alkaloids Hydrolysis of the atropine yielded tropic acid ($<0.02 \times 10^5$ dpm/mM) and tropine (8.1×10^5 dpm/mM). Reaction of the tropine with phenyl chloroformate¹⁴ afforded *N*-carbophenoxytropine (0.04×10^5 dpm/mM). The scopolamine on hydrolysis yielded tropic acid ($<0.03 \times 10^5$ dpm/mM) and oscine (8.05×10^5 dpm/mM).

RS-Phenyl-lactic-[1-¹⁴C] *acid* Glycine-[1-¹⁴C] (Amersham-Searle, nominal activity 0.5 mCi, 101 mg, 1.35 mM) was dissolved in H₂O (1 ml) and Ac₂O (1 ml) added. After stirring for 1 hr at room temp. the mixture was diluted with H₂O and extracted with pentane in a continuous extractor for 18 hr. The aq. soln. was then neutralized with NaOH and evaporated to dryness. The residual sodium acetate was dissolved in a mixture of Ac₂O (2 ml) and benzaldehyde (0.5 ml) and refluxed for 1.5 hr in N₂. The reaction mixture was cooled and diluted with H₂O (2 ml) when the azlactone of *N*-acetyl- α -amino-cinnamic acid separated (72 mg). Recrystallization from CCl₄ afforded the azlactone as yellow needles (63.1 mg) m.p. 148.5–149.5 (lit.¹⁵ m.p. 148–150°). This labeled azlactone (61.4 mg) was diluted with inactive material (52.2 mg) and refluxed in 2 N HCl (4 ml) in N₂ for 2 hr. The cooled, filtered soln. was extracted with Et₂O (4 \times 10 ml) which was then washed with satd. NaCl soln. and evaporated to dryness. The residual crude phenylpyruvic-[1-¹⁴C] acid was dissolved in a mixture of satd. Na₂CO₃ soln. (5 ml) and H₂O (2 ml), cooled to 0°, and NaBH₄ (81 mg) added. After stirring for 2 hr the soln. was acidified with HCl, and extracted with CH₂Cl₂ for 12 hr. The residue obtained on evaporation of the extract was dried by azeotrope off the H₂O with C₆H₆, and finally crystallized from CCl₄ to yield phenyl-lactic-[1-¹⁴C] acid as colorless plates (75.9 mg) m.p. 94.5–96° (lit.²¹ m.p. 94–95°) having a sp. act. of 5.15×10^8 dpm/mM (22% radiochemical yield from glycine-[1-¹⁴C]).

RS-Littorine-[1-¹⁴C] {3 α -(2-hydroxy-3-phenylpropionyloxy)-[1-¹⁴C]-tropine} Phenyl-lactic-[1-¹⁴C] acid (58 mg, 0.35 mM, 5.15×10^8 dpm/mM) was mixed with tropine (56.7 mg, 0.41 mM) and the mixture heated at 120° in a current of dry HCl gas. After 4.5 hr the mixture was cooled and the pale yellow gum stirred with 10% aq. NH₃ (10 ml). This aqueous solution was extracted with CH₂Cl₂ (4 \times 10 ml) which was then extracted with 2 N HCl (2 \times 10 ml) and H₂O (10 ml). The combined aqueous extracts were brought to pH 8 by the addition of Na₂CO₃ and extracted with Et₂O. Evaporation yielded littorine as a colorless oil (67 mg) which was converted to its picrate and crystallized from EtOH (80.3 mg) m.p. 158.5–160° (lit.¹⁶ m.p. 159.5–160°) (sp. act. 4.6×10^8 dpm/mM). Littorine-[3H] was prepared similarly from phenyllactic acid and tropine-[3-³H]¹⁴ and had a specific activity of 1.02×10^9 dpm/mM.

Administration of RS-3 α -(2-Hydroxy-3-phenylpropionyloxy)-[1-¹⁴C]-tropine-[3-³H] to *D. stramonium* and isolation of the alkaloids RS-Littorine-[1-¹⁴C] picrate (80 mg, 0.154 mM) and RS-littorine-[3H] picrate (246 mg, 0.467 mM) were dissolved in 2 N HCl (60 ml) and the soln. extracted with Et₂O until free of picric acid. The aq. soln. was evaporated to dryness *in vacuo* at 30° and the residue exposed to high vacuum to remove traces of HCl and H₂O. The residual littorine hydrochloride was dissolved in H₂O (10 ml) and assayed. The soln. contained ¹⁴C 7.05×10^7 dpm, ³H 4.76×10^8 dpm (³H/¹⁴C = 6.75). The sp. act. (¹⁴C) 1.13×10^8 dpm/mM. This aqueous solution was administered to six *D. stramonium* plants (3-months-old) as before. The plants were harvested 7 days later (fr. wt. 695 g) and the alkaloids isolated without the addition of inactive atropine. Partition chromatography of the crude alkaloids (¹⁴C 2.1×10^7 dpm, ³H/¹⁴C = 7.1) on a celite column¹ yielded a mixture of atropine and littorine which was separated by TLC on Silica gel PF-254, developing with CHCl₃-EtOH (19:1) saturated with anhydrous NH₃. The littorine (43 mg) was converted to its picrate and crystallized to constant activity ¹⁴C 1.14×10^8 dpm/mM (³H/¹⁴C = 6.9). The atropine (51.6 mg) was sublimed (95°, 5×10^{-4} mm) and crystallized to constant activity as its hydrobromide ¹⁴C 6.80×10^5 dpm/mM, ³H 2.26×10^7 dpm/mM (³H/¹⁴C = 33).

Degradation of the atropine Hydrolysis of the atropine yielded tropine assayed as its HBr salt (³H 2.02×10^7 dpm/mM, negligible ¹⁴C) and tropic acid (¹⁴C 6.82×10^5 dpm/mM, negligible ³H). The tropine dissolved in CH₂Cl₂ was oxidized with a soln. of CrO₃ in pyridine²² affording tropinone assayed as its picrate (negligible activity). The tropic acid was degraded as previously described.⁵ Dehydration afforded atropic acid (6.54×10^5 dpm/mM) which was oxidized with NaIO₄ and OsO₄ yielding formaldehyde (inactive) and phenylglyoxylic acid isolated as its oxime (6.9×10^5 dpm/mM). This oxime was decomposed in boiling H₂O yielding CO₂ collected as BaCO₃ (6.64×10^5 dpm/mM) and benzonitrile which was hydrolysed to benzoic acid (inactive).

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²¹ LIEBISCH, H. W., BHAVSAR, G. C. and SCHULTZ, H. R. (1971) *Z. Chem.* **12**, 220.

²² RATCLIFFE, R. and RODEHORST, R. (1970) *J. Org. Chem.* **35**, 4000.